

Innovations

A Comprehensive Review on Role of Salicylic Acid (SA) Against Lead (Pb) Induced Toxic Responses in Plants

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Abstract: Lead (Pb) toxicity emerging as a major environmental issue because of its pervasiveness and persistence, which negatively impacts the growth and development of agriculturally important crops. Pb exposure causes serious physiological and biochemical changes in plants, such as oxidative stress, reduced photosynthesis, changed nutrient uptake, stunted growth, impaired metabolism, etc. Although, plants have diverse inherent mechanisms, for example, activation of antioxidant enzymes, phytochelatin synthesis, osmolyte production, etc. to cope with metal toxicity, however they are not sufficient to alleviate the toxicity completely. In order to combat stress induced damages, recently application of various phytohormones and signaling molecules has drawn interest due to their potential to strengthen plant defenses against heavy metal stress. A naturally occurring phenolic compound called salicylic acid (SA) has become one of the most important modulators of plant stress tolerance. SA improves plant resistance to metal stress by modulating various physiological and biochemical reactions. Moreover, SA aids in the control of metal accumulation and transport by regulating ionic homeostasis and modulating antioxidant defense machinery. By keeping all this in mind, the present study addresses the role of SA in eliminating the negative impact of Pb toxicity on plants. Moreover, this review also highlights the interactive role of SA with other phytohormones in negating the impact of diverse abiotic stressors on plants, through sustainable manner.

Keywords: Lead, Salicylic Acid, Reactive Oxygen Species, Photosynthesis, Antioxidant, Nutrients

Introduction

Heavy metals (HMs) are the most common contaminants found in agricultural field, worldwide. Excessive accumulation of HMs not only contaminate the soil but also have an adverse effect on the safety and quality of food (Angon et al., 2024). HMs enters into the field by anthropogenic activities (e.g. industrialization,

mining, use of fertilizers, pesticides, etc.) and natural sources (such as volcanic eruption, rock weathering, etc.). HMs such as arsenic (As), Lead (Pb), cadmium (Cd) and nickel (Ni), etc. are the most prevalent metal ions detected in metal polluted areas (Subasinghe et al., 2022). The majority of HMs are non-biodegradable and therefore, with time their total concentration is building up in soils. Among the different HMs, Pb belonging to group IV of the periodic table, is a potential HM which, ranks 2nd in toxicity as per the substance priority list (ATSDR 2022). It is found naturally in the earth crust with bluish-grey appearance and have various negative effects on growth and metabolism of the plant. The major predominant forms of Pb in soil are lead oxides and hydroxides, lead-metal oxyanion complexes, Pb (II), etc. (Dongre et al., 2021). Moreover, it is also found as a combination with other elements such as oxygen (PbCO_3), sulphur (PbS , PbSO_4), etc. The range of Pb in earth's crust varies from 10 to 30 mg kg^{-1} and in the surface soils, it varies from 10 to 67 mg kg^{-1} (Frank et al., 2019).

Plants majorly take up Pb from the soil through the roots and accumulate it in different plant parts. Moreover, it can also be absorbed by the leaves and thereafter, can be transferred to other aerial parts of the plant. By entering into the plants, it leads to the negative impact on morphology, growth, and photosynthetic processes (Sharma et al., 2005). The development and growth of plants are seriously threatened by Pb toxicity. Even in small amounts, it disrupts a number of physiological and biochemical functions in plants. Growing Pb concentrations dramatically reduced seed germination, growth and increased membrane permeability in three sorghum varieties (Osman et al., 2023). Pb toxicity has also been reported to alter the structure of roots which lead to a significant reduction in growth and biomass (Fahr et al., 2013). Pb toxicity impairs the root development which make it difficult for the plant to absorb nutrients and water. Moreover, Pb toxicity reduced the root length, biomass, nutrient uptake and overall vegetative growth of *Tagetes erecta* (Gabash et al., 2023). By decreasing the permeability of cell membranes, Pb also interferes with the plant's ability to absorb and transport water (Raniet al., 2024). Furthermore, by competing for uptake and transport with vital elements including calcium, magnesium, and potassium, Pb results in nutritional imbalances. Deficiencies resulting from this competition further hinder growth and metabolism. Moreover, Pb poisoning increases the generation of reactive oxygen species (ROS), including hydrogen peroxide and superoxide radicals, which leads to oxidative damage to the plants (Mansoor et al., 2023). Various cellular processes are compromised by these ROS because they directly harm proteins, lipids, and nucleic acids. Despite the fact that plants use antioxidant defense mechanisms to fight oxidative stress, extended exposure to Pb cannot overcome the damage caused by ROS and therefore, reduce the growth and productivity of crop plants and leads to cellular damage. Nowadays, SA a member of the remarkably varied class of plant phenolics, is crucial for controlling plant growth and development of plants. SA reduces the vulnerability of plants to environmental stressors by regulating the photosynthetic

rate, transpiration rates, stomatal movement, and antioxidant defense system. SA increases the net CO₂ assimilation rate by improving shoot dry matter (Nazar et al., 2015). Moreover SA reported to enhance the photosynthetic pigments, Osmo protection, membrane stability, carbon metabolism, and antioxidant system in wheat, barley, rice, maize, and tomato (Das et al., 2024). The beneficial impacts of SA on plants' reactions to abiotic stressors, such as ozone and ultraviolet (UV) radiation heat stress, cold and drought, and salt and osmotic stresses have been shown in several studies (Yang et al., 2023). Interestingly, SA plays a crucial role in the plant's immune system under biotic stress circumstances, especially through systemic acquired resistance (SAR). Although, various studies have been conducted in reviewing the impact of SA in plants under diverse abiotic stresses, however, no review is available on unravelling the role of SA under Pb toxicity in different plant species. Therefore, current study will provide an overview on impact of SA in imparting stress tolerance to the plants by modulating various physiological, biological and molecular processes.

Effect of lead (Pb) on plants

Seed germination and growth: Plants majorly take up Pb from the soil through the roots and accumulate it in different plant parts. Moreover, it can also be absorbed by the leaves and thereafter, can be transferred to other aerial parts of the plant. By entering into the plants, it leads to the negative impact on morphology, seed germination, growth, and photosynthetic processes. For example, Awan et al. (2015) reported reduced germination index, germination percentage, and plant biomass of rice under Pb toxicity. Similarly, Pb toxicity resulted in shortening of roots and hypocotyl of Lupinus plants and thereby reduced the number of germinating seeds (Wozny et al., 1982). Pb toxicity resulted in reduced root and shoot biomass of cow pea plants due to higher accretion of Pb in respective tissues (Kopittke et al., 2007). Jiang and Liu (2010) correlated the reduced growth of Pb exposed *Allium sativum* with the alteration in ultrastructure of endoplasmic reticulum and disruption of biological membranes. Zea mays plants displayed reduction in germination percentage, seedling growth, and protein content under Pb stress due to inhibited activities of various vital enzymes (amylase, protease etc) (Hayat et al., 2010).

Similarly in rice endosperm, reported reduced amylase and protease activity under Pb toxicity which led to reduction in seed germination rate (Wang et al., 2020). Exposure of *Spartiana alterniflora* and *Pinus helipensis* seeds with Pb displayed reduced germination percentage (Staszak et al., 2020). Pb toxicity delayed the germination trait and reduced the germination ability of seeds in different species of families Cruciferae, papilionaceae and poaceae. Soil contaminated with Pb toxicity displayed reduced seed germination rate and undesirable morphological traits in *Ocimum basilicum* (Fattahi et al., 2019). Pb toxicity inhibited the growth of roots and shoots in Zea mays, *Triticum aestivum*, and *Medicago sativa* in concentration dependent manner (Vasilachi-Mitoseru

etal., 2023). On the same line, exposure of wheat plants with Pb toxicity led to a significant reduction in plant biomass, height and root and shoot growth with greater negative impact on root biomass than the shoots (Kumar et al., 2018). Talha et al. (2023) correlated the negative impact on root growth of maize seedling with increasing dosage of Pb in the rooting media. In the study conducted by Chandrasekhar and Ray (2019) rising lead nitrate levels in the soil had a detrimental effect on the biomass of the roots and shoots of *Eclipta prostrata*, *Scoparia dulcis*, and *Phyllanthus niruri* with lesser impact of Pb stress on *Eclipta prostrata* than the other two species. Overall, the studies highlight that the application of heavy metals including Pb reduced the seed germination, seedling length, growth and overall development of plants in a concentration dependent manner.

Photosynthesis, respiration and nutrient uptake: One of the most important physiological characteristics of plants is photosynthesis, which plays a vital role in growth and productivity of plants. However presence of HM in the rooting medium have detrimental impact on the photosynthetic machinery of plants as well as its functions, and thereby can reducing the chlorophyll synthesis and inhibit the Calvin cycle either directly or indirectly through blocking the light and dark reactions (Muhammad et al., 2021). When sugar beet plants were exposed with varying concentration of Pb, a significant decline in growth and photosynthetic efficacy was observed by Wu et al. (2021). Various studies have examined the effects of Pb toxicity on chlorophyll fluorescence, growth, photosynthetic pigments, and found adverse effect of the heavy metal on these aspects. For example Ahmad et al. (2008) reported that Pb inhibited the C₃ cycle and the electron transport chain, and thereby, prevented the synthesis of vital pigments through altering the ultrastructure of chloroplasts. Additionally, Cencki et al. (2010) reported interference of Pb with other nutrients uptake (Fe, Mn, S) etc. which led to inhibition of photosynthesis in *Brassica rapa*. Pb toxicity inhibited the activity of enzymes involved in nitrate assimilation that is nitrate reductase enzyme, which decreased the nitrogen content in *Brassica pekinensis* (Xiong et al., 2006). In a similar way, high Pb concentrations decreased the amount of phosphorous (P) in cabbage, which in turn reduced plant growth and biomass accumulation (Sinha et al., 2006). Pb stress affected mitochondrial activity in *S. drummondii* leading to reduced cellular respiration rate (Ruley et al., 2006). Kibria et al. (2009) revealed the negative impact of Pb on nutritional status, growth and germination of *Amaranthus gangeticus* and *A. oleracea*. Moreover, Pb poisoning interfered with the absorption of nutrients by competing with necessary components, resulting in insufficiencies in iron (Fe), calcium (Ca), and magnesium (Mg). Photosynthetic efficiency of *Phaseolus vulgaris* was negatively impacted by Pb poisoning, due to decreased photosynthetic pigments, nutrient uptake and limited light absorption (Khalil et al., 2021). Exposure of *Leucaena leucocephala* to Pb toxicity resulted in alteration of ultrastructure of the

chloroplasts and lowering of nutrient uptake which ultimately led to dramatic decrease in the photosynthetic rate (Alkhatib et al., 2019). According to the study of Gupta et al. (2024), Pb poisoning in *Zea mays* and *Oryza sativa* hampered photosynthesis by destroying chloroplast structures, altering nutrient uptake (Mg, K, N) interfering stomatal conductance and breaking down chlorophyll pigments. Algethami et al. (2023) reported that Pb and Cd stress in *Triticum aestivum* reduced chlorophyll content, hampered gas exchange parameter and nutrient acquisition (N, P, K) which had a detrimental effect on photosynthetic efficiency of plants.

Oxidative stress and antioxidant defense: Lipids and proteins in plant cells are the main targets of oxidative damage brought on by ROS (Anjum et al., 2015). For example, *Citrus aurantium* L. plants grown under Pb enriched soil experienced higher oxidative stress due to increased production of ROS (Giannakoula et al., 2021). Similarly, Sun et al. (2010) reported higher accumulation of H_2O_2 concentrations in roots of *Hypnum plumaeforme* subjected to Pb stress. Pb toxicity increased the production of H_2O_2 in roots of *B. juncea* which led to oxidative damage to the plants (Dalyan et al., 2018). Similarly, Huang et al. (2008) reported higher burst of ROS in two different ecotypes of *Sedum alferdii* under Pb toxicity. Increased ROS and MDA levels were reported by Wang et al. (2012) in *Vallisneria spiralis* when exposed to Pb indicating higher level of oxidative stress in metal stressed plant when compared to control. On the same line, elevated level of ROS were reported in Pb stressed *Coronopus didymus* further indicating the role metal toxicity in inducing oxidative burden on plants (Sidhu et al., 2016). Plants have a variety of defense mechanisms that work to eliminate ROS before they have a chance to harm delicate components of the cellular machinery. These mechanisms can be easily separated into two categories: enzymatic antioxidants (catalase, superoxide dismutase, glutathione reductase) and non-enzymic antioxidants (glutathione, ascorbate, carotenoids) etc, which have the ability to nullify the impact of oxidative stress. The antioxidative enzymes GR, GPOX, superoxide dismutase etc. were found to be more upregulated in the roots than leaves of rice plants under Pb toxicity (Verma and Dubey, 2003). Pb toxicity can cause a variety of symptoms, including impaired mitotic division and altered activities of antioxidant enzymes, even at the cellular level (Kang et al., 2009; Kaur et al., 2012). According to Bhatti et al. (2013) under Pb metal stress, levels of chlorophyll a and chlorophyll b significantly decreased in different varieties of wheat. According to Sharma et al. (2023), Pb toxicity in barley raised oxidative stress by increasing lipid peroxidation and ROS levels and decreasing the activity of antioxidative enzymes; SOD, POD, CAT. Similarly, higher production of ROS were reported in *Brassica juncea* and *Oryza sativa* under Pb exposure due to alteration of activity of various antioxidant enzymes which resulted in physiological and cellular damage (Mitra et al., 2020). Khan et al. (2021) reported a significant increase in the production of ROS (MDA, H_2O_2 , etc.) and decrease in

the activity of antioxidant enzymes such as (CAT) and (SOD) when rice plants were treated with Pb. Overall, Pb toxicity has a negative impact on plant health by impairing Photosynthesis, impeding nutrient uptake, respiration, disrupting the oxidative stress and antioxidant defense system, ultimately compromising overall plant growth and productivity.

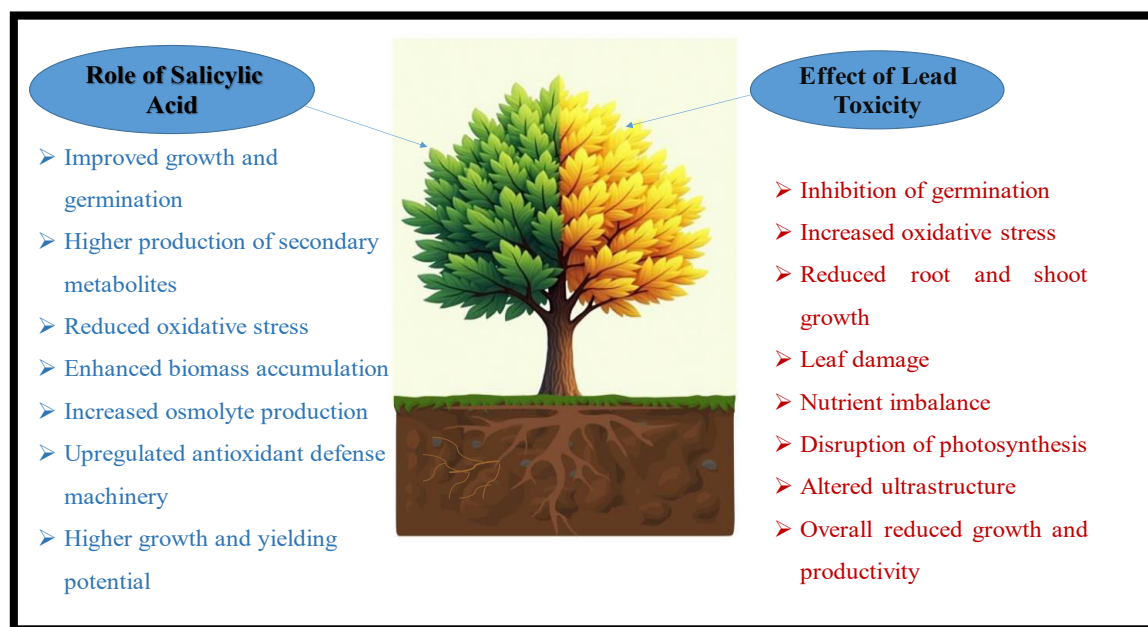


Fig 1:- A schematic presentation of role of SA in alleviating the impact of pb toxicity on plants

Role of SA in mitigation of Pb toxicity

Salicylic acid (SA), is a potential phytohormone that helps the plants to grow under unfavourable environmental conditions (Arif et al., 2020). It regulates various physiological, biochemical and molecular processes which, defend the plants against various stressors (Zulfiqar et al., 2021) (Fig 1). SA has been reported to reduce the effects of various HMs on a variety of crops (Table 1). For example, application of SA improved the root and shoot length of *Oryza sativa* seedlings when exposed to Pb stress (Jing et al., 2007). Similarly, SA pretreatment enhanced the biomass production, shoot and root growth, and chlorophyll levels in maize plants when exposed to Pb toxicity (Zanganeh et al., 2020). In *Brassica napus*, SA increased biomass accumulation, preserved water balance, and decreased Pb toxicity by boosting the seedling growth (Jazi et al., 2015). Fontenele et al. (2017) observed a significant improvement in plant height, leaf number, and dry weight when *Vigna unguiculata* were treated with SA.

In addition to growth attributes, SA can improve the rubisco activity, stabilize the structure of chloroplast, increased the accumulation of photosynthetic pigments etc. under stressful conditions (Mateo et al., 2006, Hayat et al., 2007). Moreover, SA plays a crucial role in nutrient acquisition, water uptake and maintaining the integrity of the cellular membrane (Khokon et al., 2011). For instance, amendment

of maize plants with SA displayed higher photosynthetic efficiency, nutrient uptake and reduced Pb accumulation (Gupta et al., 2024). In *Vigna unguiculata* under Pb stress, exogenous supplementation of SA improved the photosynthetic pigments level, stabilised nutrient intake, and lessened the impact of oxidative stress (Fontenele et al., 2017). Similarly, SA reduced the intensity of oxidative stress by improving the nutritional status (Mn, Ca, Fe) and inhibited the accumulation of Pb in *Vallisneria natans* (Wang et al., 2011).

Besides, SA decreased the damage caused by oxidative species by increasing the transcription of antioxidant defense genes such as POD, CAT, SOD in *Brassica campestris* when exposed to Pb stress (Hasanuzzaman et al., 2019). *Brassica juncea* under Pb stress showed increased activity of antioxidant enzymes, including APX, CAT and SOD (Agnihotri et al., 2018). SA treatment reduced the intensity of oxidative stress by improving the antioxidant enzymes activity in Pb stressed *Brassica napus* (Sharma et al., 2020). When Pb stressed wheat plants were supplied with SA, it accelerated the antioxidant defense machinery of stressed plant and prevented them from damage caused by metal toxicity (Chen et al., 2016). SA pre-treatment reduced the generation of oxidative species and increased antioxidant enzymes activity which in turn reduced the oxidative damage to *Allium cepa* (Kaur et al., 2021). Similarly, SA increased the synthesis of phenolic compounds, organic acids and modulated antioxidant enzymes activity and thereby reduced the effect of oxidative damage in *Brassica juncea* exposed to Pb stress (Kholi et al., 2017). On the same way, SA reduced the accumulation of ROS and enhanced the glutathione (GSH) levels as well as the activities of antioxidant enzymes including CAT in Pb-stressed *Zea mays* plants (Gupta et al., 2024). On the same line, exogenous supplementation of SA reduced the production of ROS in Pb stressed *B. campestris* plants by upregulating the antioxidant defense machinery and thereby, enhanced the growth and productivity (Hasanuzzaman et al., 2019).

SA is well known to enhance the accumulation of osmolytes (proline, glycine betaine, and soluble sugars), which aid in preserving osmotic balance and maintaining cellular structures. For example, SA application under Pb stress inclined the synthesis of osmolytes (proline, sugar) which imparted metal tolerance to *Brassica juncea* (Kohli et al., 2018). Exogenous application of SA increased the osmolyte synthesis (proline and soluble sugars) in Pb stressed Faba bean and common bean which helped the plants to maintain cellular homeostasis and thereby mitigated the negative effects of metal stress (Layachi et al., 2023). Similarly, exogenous application of SA to Pb stressed maize plants displayed improved tolerance towards metal toxicity by reducing the level of methylglyoxal, increasing the NO content in shoots and roots, and preserving the balance of cysteine and methionine (Zanganeh et al., 2019). Therefore, regulation of ROS detoxification machinery to shield plants from metal-induced damages is another important mechanism underlying SA-regulated metal tolerance in plants (Saleem et al., 2021, Song et al., 2023). All the above studies highlight that SA can

activate plant defense mechanisms by upregulating the activity of antioxidant enzymes, maintaining cellular redox homeostasis, and altering the level of transcripts encoding various stress responsive pathways.

Table 1:-Mode of action of SA under heavy metal stress

Sr No.	Host plant	Concentration of salicylic acid	Mode of action of salicylic acid	References
1.	Lycopersicon esculentum	1 mM	Reduced oxidative stress markers, Na level, increased nutrient uptake (Mg, K) and upregulated antioxidant defense machinery	He and Zhu (2008)
2.	Daucus carota	0, 0.5 mmol kg ⁻¹	Improved plant biomass, photosynthetic pigment concentration (carotenoids, anthocyanin) and upregulated antioxidant enzymes activity	Eraslan et al. (2007)
3.	Ctenanthes esculenta	10 ⁻⁶ M	Decreased water loss and upregulated the antioxidant defense mechanism	Kadioglu et al. (2011)
4.	Citrullus lanatus	0, 0.25, 0.50, 0.75 and 1.0 µmol L ⁻¹	Increased osmolyte production and reduced membrane damage	Silva et al. (2023)
5.	Helianthus annuus L.	0, 100, 200, 300 mg L ⁻¹	Increased leaf SOD and POD activity, growth and photosynthetic capacity	Noreen et al. (2009)
6.	Satureja khuzistanica	100 and 200 mg L ⁻¹	Increased growth, essential oil content and yield	Sadeghian et al. (2013)
7.	Lycopersicon esculentum	10 ⁻⁷ and 10 ⁻⁴ M	Reduced oxidative damage and increased antioxidant enzymes activity	Szepesi (2008)
8.	Triticum aestivum	1, 2, 4, 8, and 16 mg L ⁻¹	Increased growth, chlorophyll content and activities of various antioxidant enzymes	Wang and Zhang (2017)
9.	Cicer arietinum	0.5 mM	Increased biomass, productivity, higher ROS	Kaur et al. (2022)

	um		scavenging, improved nodulation potential and photosynthetic efficiency	
10.	Medicago sativa	0.1 and 0.5 mM	Increased plant growth and photosynthetic capacity, inhibited catalase activity and prevented the accumulation of polyamines.	Palma et al. (2013)
11.	Vigna radiata	0.5 mM	Increased glycine betaine (GB) accumulation suppressed ethylene formation and enhanced antioxidant defense responses and photosynthetic efficacy	Khan et al. (2014)
12.	Pisum sativum L.	1.0 mM	Increased SOD, APX, CAT activities and lesser production of ROS	Singh et al. (2015)
13.	Hordeum vulgare	500 μ M	Enhanced antioxidant defense abilities and photosynthetic efficiency	Habibi (2012)
14.	Pisum sativum L.	0.1 mM	Reduced Pb-induced oxidative stress by improving growth characteristics, photosynthetic pigment contents, and antioxidant enzyme activities	Popova et al. (2011)
15.	Calendula officinalis	0, 1, 2 mM	Increased plant biomass, height, leaf area, no. of flowers and chlorophyll content	Bayat and Neamati (2012)

Crosstalk between SA with phytohormone

SA is a versatile phytohormone that plays a key role in growth and development of plants by coordinating multiple defense responses under unfavourable environmental conditions. The conversion points in signal transduction pathways of hormones, have a significant role in regulating the growth and stress responses in plants. Coordination between SA and other phytohormones (auxins, cytokinin, gibberellins, ABA, ethylene, and jasmonates) is a crucial factor that regulate diverse stress responses in plants facing multiple challenges (Nishiyama et al., 2013). Therefore, it is necessary to comprehend the intricate relationship between pathways of different phytohormone. SA treatment reduced the negative effects of Pb stress on pepper plants by improving the endogenous H₂S content and

modulating ascorbate glutathione pool (Kaya et al., 2023). Moreover, by upregulating the activity of enzymes involved in H₂S biosynthesis, the combined application of SA and H₂S imparted heat tolerance to maize plant (Li et al., 2015). Individual and cumulative application of SA and kinetin improved various physiological and biochemical parameters in Cd-stressed wheat plants (Eydi Asl Shoshtari et al., 2024). On the same line, Cd stressed wheat seedling treated with SA and indole-3-acetic acid displayed synergistic effects of both the phytohormones in term of improved antioxidant enzymes activity and altered ultrastructure of leaf (Agami et al., 2013). Exogenous application of SA reduced the level of ethylene and thereby safeguard the rice plants from detrimental impact of As (Mahajan et al., 2023). A study conducted by Kohli et al. (2018) displayed that individual as well as cumulative application of SA and 24-epibrassinolide decreased the damaging effects of Pb on *Brassica juncea* by improving photosynthetic efficiency, nutrient uptake and upregulating the activity of antioxidant enzymes. Additionally, SA and nitric oxide (NO) mitigated the harmful effects of Cd on ryegrass by improving the chlorophyll content, antioxidant enzymes activity and reducing the ROS production (Wang et al., 2013). Najafi Kakavand et al., (2018) reported increased osmolytes and carotene accumulation, antioxidant enzymes activity when Ni-stressed *Alyssum inflatum* were treated with SA and jasmonic acid (JA). Similarly, SA and gibberellic acid reduced metal induced oxidative damage in three different cultivars of *Vigna unguiculata* (Ahmad et al., 2021). Overall, the studies highlight that coordination between SA signaling and other phytohormones is a crucial factor to mitigate the negative impact of metal stress on various plant species.

Conclusions

Pb toxicity significantly hinders the growth and development by disrupting various physiological and metabolic processes in plants. Its buildup in plant tissues endangers food safety in addition to influencing biomass accumulation. Nowadays, application of SA is emerging as a promising approach for increasing the resilience of crop plants, including under metal toxicity. SA has a variety of protective properties, such as the capacity to control physiological functions, modify antioxidant defense systems, and reduce oxidative damage brought on by ROS generated by metal toxicity. Additionally, SA increases nutrient uptake, stabilizes membrane integrity, increases photosynthetic efficiency, and preserves chlorophyll content all of which are essential for sustaining plant vitality. Interestingly, the cumulative application of SA and other phytohormone also proved as an innovative approach in alleviating the detrimental impact of various abiotic stressors. However, the specific report on individual as well as interactive role of SA in negating the effect of Pb stress on agriculturally important crop is scanty and needs further attention. Moreover, very few reports are available on the precise molecular mechanisms underlying SA-

mediated detoxification processes, which could be a major area of research for the scientists working in the same field.

References

1. Agami, R. A., and Mohamed, G. F. (2013). Exogenous treatment with indole-3-acetic acid and salicylic acid alleviates cadmium toxicity in wheat seedlings. *Ecotoxicology and environmental safety*, 94, 164-171.
2. Agnihotri, A., Gupta, P., Dwivedi, A., and Seth, C. S. (2018). Counteractive mechanism (s) of salicylic acid in response to lead toxicity in *Brassica juncea* (L.) Czern. cv. Varuna. *Planta*, 248, 49-68.
3. Ahmad, M. S. A., Hussain, M. U. M. T. A. Z., Ijaz, S. A. M. I. N. A., and Alvi, A. K. (2008). Photosynthetic performance of two mung bean (*Vigna radiata*) cultivars under lead and copper stress. *Int. J. Agric. Biol*, 10, 167-172.
4. Ahmad, P., Raja, V., Ashraf, M., Wijaya, L., Bajguz, A., and Alyemeni, M. N. (2021). Jasmonic acid (JA) and gibberellic acid (GA3) mitigated Cd-toxicity in chickpea plants through restricted cd uptake and oxidative stress management. *Scientific Reports*, 11(1), 19768.
5. Algethami, J. S., Irshad, M. K., Javed, W., Alhamami, M. A., and Ibrahim, M. (2023). Iron-modified biochar improves plant physiology, soil nutritional status and mitigates Pb and Cd-hazard in wheat (*Triticum aestivum* L.). *Frontiers in Plant Science*, 14, 1221434.
6. Alkhatib, R., Mheidat, M., Abdo, N., Tadros, M., Al-Eitan, L., and Al-Hadid, K. (2019). Effect of lead on the physiological, biochemical and ultrastructural properties of *Leucaena leucocephala*. *Plant Biology*, 21(6), 1132-1139.
7. Angon, P. B., Islam, M. S., Das, A., Anjum, N., Poudel, A., and Suchi, S. A. (2024). Sources, effects and present perspectives of heavy metals contamination: Soil, plants and human food chain. *Heliyon*, 10(7).
8. Anjum, N. A., Sofo, A., Scopa, A., Roychoudhury, A., Gill, S. S., Iqbal, M., and Ahmad, I. (2015). Lipids and proteins—major targets of oxidative modifications in abiotic stressed plants. *Environmental Science and Pollution Research*, 22, 4099-4121.
9. Arif, Y., Sami, F., Siddiqui, H., Bajguz, A., and Hayat, S. (2020). Salicylic acid in relation to other phytohormones in plant: A study towards physiology and signal transduction under challenging environment. *Environmental and Experimental Botany*, 175, 104040.
10. Awan, S., Jabeen, M., Imran, Q. M., Ullah, F., Mehmood, Z., Jahngir, M., and Jamil, M. (2015). Effects of lead toxicity on plant growth and biochemical attributes of different rice (*Oryza Sativa* L.) varieties. *Journal of Bio-Molecular Sciences*, 3(1), 44-55.
11. Bayat, H., Alirezaie, M., and Neamati, H. (2012). Impact of exogenous salicylic acid on growth and ornamental characteristics of calendula (*Calendula officinalis* L.) under salinity stress. *Journal of Stress Physiology and Biochemistry*, 8(1), 258-267.

12. Bhatti, K. H., Sehrish Anwar, S. A., Khalid Nawaz, K. N., Khalid Hussain, K. H., Siddiqi, E. H., Sharif, R. U., and Aneela Khalid, A. K. (2013). Effect of heavy metal lead (Pb) stress of different concentration on wheat (*Triticum aestivum* L.).
13. Cenkci, S., Ciğerci, İ. H., Yıldız, M., Özay, C., Bozdağ, A., and Terzi, H. (2010). Lead contamination reduces chlorophyll biosynthesis and genomic template stability in *Brassica rapa* L. *Environmental and experimental botany*, 67(3), 467-473.
14. Chandrasekhar, C., and Ray, J. G. (2019). Lead accumulation, growth responses and biochemical changes of three plant species exposed to soil amended with different concentrations of lead nitrate. *Ecotoxicology and Environmental Safety*, 171, 26-36.
15. Chen, Y. E., Cui, J. M., Li, G. X., Yuan, M., Zhang, Z. W., Yuan, S., and Zhang, H. Y. (2016). Effect of salicylic acid on the antioxidant system and photosystem II in wheat seedlings. *Biologia plantarum*, 60(1), 139-147.
16. Dalyan, E., Yüzbaşıoğlu, E., and Akpınar, I. (2018). Effect of 24-epibrassinolide on antioxidative defence system against lead-induced oxidative stress in the roots of *Brassica juncea* L. seedlings. *Russian Journal of Plant Physiology*, 65, 570-578.
17. Das, A. K., Ghosh, P. K., Nihad, S. A. I., Sultana, S., Keya, S. S., Rahman, M. A., and Rahman, M. M. (2024). Salicylic Acid Priming Improves Cotton Seedling Heat Tolerance through Photosynthetic Pigment Preservation, Enhanced Antioxidant Activity, and Osmoprotectant Levels. *Plants*, 13(12), 1639.
18. Dongre, R. S. (2021). Chromium and lead as soil pollutants: insights on toxicity profiles and their remediation. *Journal of Advanced Biotechnology and Bioengineering*, 9, 1-16.
19. Eraslan, F., Inal, A., Gunes, A., and Alpaslan, M. (2007). Impact of exogenous salicylic acid on the growth, antioxidant activity and physiology of carrot plants subjected to combined salinity and boron toxicity. *Scientia horticulturae*, 113(2), 120-128.
20. Eydi Asl Shoshtari, B., Rahnama, A., Hassibi, P., and Zoufan, P. (2024). Effects of salicylic acid and kinetin on some physiological, biochemical traits and the accumulation of cadmium in durum wheat. *Journal of Plant Biological Sciences*, 16(1), 39-59.
21. Fahr, M., Laplaze, L., Bendaou, N., Hoher, V., Mzibri, M. E., Bogusz, D., and Smouni, A. (2013). Effect of lead on root growth. *Frontiers in plant science*, 4, 175.
22. Fattahi, B., Arzani, K., Souri, M. K., and Barzegar, M. (2019). Effects of cadmium and lead on seed germination, morphological traits, and essential oil composition of sweet basil (*Ocimum basilicum* L.). *Industrial Crops and Products*, 138, 111584.
23. Fontenele, N. M. B., Otoch, M. D. L. O., Gomes-Rochette, N. F., de Menezes Sobreira, A. C., Barreto, A. A. G. C., de Oliveira, F. D. B., and de Melo, D. F. (2017).

- Effect of lead on physiological and antioxidant responses in two Vigna unguiculata cultivars differing in Pb-accumulation. Chemosphere, 176, 397-404.*
24. Frank, J. J., Poulakos, A. G., Tornero-Velez, R., and Xue, J. (2019). Systematic review and meta-analyses of lead (Pb) concentrations in environmental media (soil, dust, water, food, and air) reported in the United States from 1996 to 2016. *Science of the Total Environment*, 694, 133489.
 25. Gabash, H. M., Resan, A. Z., and Jasim, F. M. (2023). The Effect Of Different Levels Of Lead On The Vegetative And Root Growth Of Tagetes Erect L. *British Journal of Global Ecology and Sustainable Development*, 16.
 26. Giannakoula, A., Therios, I., and Chatzissavvidis, C. (2021). Effect of lead and copper on photosynthetic apparatus in citrus (*Citrus aurantium* L.) plants. The role of antioxidants in oxidative damage as a response to heavy metal stress. *Plants*, 10(1), 155.
 27. Gupta, M., Dwivedi, V., Kumar, S., Patel, A., Niazi, P., and Yadav, V. K. (2024). Lead toxicity in plants: mechanistic insights into toxicity, physiological responses of plants and mitigation strategies. *Plant Signaling and Behavior*, 19(1), 2365576.
 28. Gupta, M., Kumar, S., Dwivedi, V., Gupta, D. G., Ali, D., Alarifi, S., and Yadav, V. K. (2024). Selective synergistic effects of oxalic acid and salicylic acid in enhancing amino acid levels and alleviating lead stress in *Zea mays* L. *Plant Signaling and Behavior*, 19(1), 2400451.
 29. Habibi, G. (2012). Exogenous salicylic acid alleviates oxidative damage of barley plants under drought stress. *Acta Biologica Szegediensis*, 56(1), 57-63.
 30. Hasanuzzaman, M., Matin, M. A., Fardus, J., Hasanuzzaman, M. D., Hossain, M. S., and Parvin, K. (2019). Foliar application of salicylic acid improves growth and yield attributes by upregulating the antioxidant defense system in *Brassica campestris* plants grown in lead-amended soils. *Acta Agrobotanica*, 72(2).
 31. Hayat, Q., Hayat, S., Irfan, M., and Ahmad, A. (2010). Effect of exogenous salicylic acid under changing environment: a review. *Environmental and experimental botany*, 68(1), 14-25.
 32. Hayat, S., Ali, B., and Ahmad, A. (2007). Salicylic acid: biosynthesis, metabolism and physiological role in plants. *Salicylic acid: A plant hormone*, 1-14.
 33. He, Y., and Zhu, Z. J. (2008). Exogenous salicylic acid alleviates NaCl toxicity and increases antioxidative enzyme activity in *Lycopersicon esculentum*. *Biologia Plantarum*, 52, 792-795.
 34. Huang, H., Li, T., Tian, S., Gupta, D. K., Zhang, X., and Yang, X. E. (2008). Role of EDTA in alleviating lead toxicity in accumulator species of *Sedum alfredii* H. *Bioresource Technology*, 99(14), 6088-6096.
 35. Jazi, S., and Oregani, K. (2015). The Role of Salicylic Acid Treatment on the Growth, Photosynthetic Pigment of *Brassica Napus* L. Under Lead Stress. *Ecology, Environment and Conservation*, 21(1), 1-8.
 36. Jiang, W., and Liu, D. (2010). Pb-induced cellular defense system in the root meristematic cells of *Allium sativum* L. *BMC Plant Biology*, 10, 1-8.

37. Jing, C. H. E. N., Cheng, Z. H. U., Li-ping, L. I., Zhong-yang, S. U. N., and Xue-bo, P. A. N. (2007). Effects of exogenous salicylic acid on growth and H₂O₂-metabolizing enzymes in rice seedlings under lead stress. *Journal of Environmental sciences*, 19(1), 44-49.
38. Kadioglu, A., Saruhan, N., Sağlam, A., Terzi, R., and Acet, T. (2011). Exogenous salicylic acid alleviates effects of long term drought stress and delays leaf rolling by inducing antioxidant system. *Plant Growth Regulation*, 64, 27-37.
39. Kang JiLi, K. J., Zeng ZhiJun, Z. Z., and Liu YuPei, L. Y. (2009). Effects of lead (Pb²⁺) stress on seed germination and seedling growth of wheat.
40. Kaur, G., Sharma, P., Rathee, S., Singh, H. P., Batish, D. R., and Kohli, R. K. (2021). Salicylic acid pre-treatment modulates Pb²⁺-induced DNA damage vis-à-vis oxidative stress in *Allium cepa* roots. *Environmental Science and Pollution Research*, 28(37), 51989-52000.
41. Kaur, G., Singh, H. P., Batish, D. R., and Kumar, R. K. (2012). Growth, photosynthetic activity and oxidative stress in wheat (*Triticum aestivum*) after exposure of lead to soil. *Journal of environmental biology*, 33(2), 265.
42. Kaur, H., Hussain, S. J., Kaur, G., Poor, P., Alamri, S., Siddiqui, M. H., and Khan, M. I. R. (2022). Salicylic acid improves nitrogen fixation, growth, yield and antioxidant defence mechanisms in chickpea genotypes under salt stress. *Journal of Plant Growth Regulation*, 41(5), 2034-2047
43. Kaya, C., Ugurlar, F., Ashraf, M., and Ahmad, P. (2023). Salicylic acid interacts with other plant growth regulators and signal molecules in response to stressful environments in plants. *Plant Physiology and Biochemistry*, 196, 431-443.
44. Khalil, R., Haroun, S., Bassyoini, F., Nagah, A., and Yusuf, M. (2021). Salicylic acid in combination with kinetin or calcium ameliorates heavy metal stress in *Phaseolus vulgaris* plant. *Journal of Agriculture and Food Research*, 5, 100182.
45. Khan, M. I. R., Asgher, M., and Khan, N. A. (2014). Alleviation of salt-induced photosynthesis and growth inhibition by salicylic acid involves glycinebetaine and ethylene in mungbean (*Vigna radiata* L.). *Plant Physiology and Biochemistry*, 80, 67-74.
46. Khan, M., Rolly, N. K., Al Azzawi, T. N. I., Imran, M., Mun, B. G., Lee, I. J., and Yun, B. W. (2021). Lead (Pb)-induced oxidative stress alters the morphological and physio-biochemical properties of rice (*Oryza sativa* L.). *Agronomy*, 11(3), 409.
47. Khokon, M. A. R., Okuma, E. I. J. I., Hossain, M. A., Munemasa, S., Uraji, M., Nakamura, Y., and Murata, Y. (2011). Involvement of extracellular oxidative burst in salicylic acid-induced stomatal closure in *Arabidopsis*. *Plant, cell and environment*, 34(3), 434-443.
48. Kibria, M. G., Islam, M., and Osman, K. T. (2009). Effects of lead on growth and mineral nutrition of *Amaranthus gangeticus* L. and *Amaranthus oleracea* L. *Soil Environ*, 28(1), 1-6.
49. Kohli, S. K., Handa, N., Sharma, A., Gautam, V., Arora, S., Bhardwaj, R., and Ahmad, P. (2018). Combined effect of 24-epibrassinolide and salicylic acid

- mitigates lead (Pb) toxicity by modulating various metabolites in *Brassica juncea* L. seedlings. *Protoplasma*, 255, 11-24.
50. Kohli, S. K., Handa, N., Sharma, A., Gautam, V., Arora, S., Bhardwaj, R., and Ahmad, P. (2018). Combined effect of 24-epibrassinolide and salicylic acid mitigates lead (Pb) toxicity by modulating various metabolites in *Brassica juncea* L. seedlings. *Protoplasma*, 255, 11-24.
 51. Kohli, S. K., Handa, N., Sharma, A., Kumar, V., Kaur, P., and Bhardwaj, R. (2017). Synergistic effect of 24-epibrassinolide and salicylic acid on photosynthetic efficiency and gene expression in *Brassica juncea* L. under Pb stress. *Turkish Journal of Biology*, 41(6), 943-953.
 52. Kopittke, P. M., Asher, C. J., Kopittke, R. A., and Menzies, N. W. (2007). Toxic effects of Pb²⁺ on growth of cowpea (*Vigna unguiculata*). *Environmental pollution*, 150(2), 280-287.
 53. Kumar, S., Sharma, P., Misra, M., and Narayan Misra, A. (2018). Lead induced root and shoot growth reduction in wheat (*Triticum aestivum* L.) is due to increase in membrane lipid peroxidation. *J. Pharmacogn. Phytochem*, 7, 2080-2083.
 54. Layachi, N., and Kechrid, Z. (2023). The Benefit Effect Of Salicylic Acid On Physio-Biochemical Characters Of Faba Bean (*Vicia faba* L.) Under Lead Stress. *Feb-Fresenius Environmental Bulletin*, 3387.
 55. Li, Z. G., Xie, L. R., and Li, X. J. (2015). Hydrogen sulfide acts as a downstream signal molecule in salicylic acid-induced heat tolerance in maize (*Zea mays* L.) seedlings. *Journal of plant physiology*, 177, 121-127.
 56. Mahajan, M., Nazir, F., Jahan, B., Siddiqui, M. H., Iqbal, N., and Khan, M. I. R. (2023). Salicylic acid mitigates arsenic stress in Rice (*Oryza sativa*) via Modulation of Nitrogen–Sulfur Assimilation, Ethylene Biosynthesis, and Defense systems. *Agriculture*, 13(7), 1293.
 57. Mansoor, S., Ali, A., Kour, N., Bornhorst, J., AlHarbi, K., Rinklebe, J., and Chung, Y. S. (2023). Heavy metal induced oxidative stress mitigation and ROS scavenging in plants. *Plants*, 12(16), 3003.
 58. Mateo, A., Funck, D., Mühlenbock, P., Kular, B., Mullineaux, P. M., and Karpinski, S. (2006). Controlled levels of salicylic acid are required for optimal photosynthesis and redox homeostasis. *Journal of Experimental Botany*, 57(8), 1795-1807.
 59. Mitra, A., Chatterjee, S., Voronina, A. V., Walther, C., and Gupta, D. K. (2020). Lead toxicity in plants: a review. *Lead in Plants and the Environment*, 99-116.
 60. Muhammad, I., Shalmani, A., Ali, M., Yang, Q. H., Ahmad, H., and Li, F. B. (2021). Mechanisms regulating the dynamics of photosynthesis under abiotic stresses. *Frontiers in plant science*, 11, 615942.
 61. Najafi-Kakavand, S., Karimi, N., Ghasempour, H. R., Raza, A., Chaichi, M., and Modarresi, M. (2023). Role of jasmonic and salicylic acid on enzymatic changes in the root of two *Alyssum inflatum* Náyr. populations exposed to nickel toxicity. *Journal of Plant Growth Regulation*, 42(3), 1647-1664.

62. Nazar, R., Umar, S., Khan, N. A., and Sareer, O. (2015). Salicylic acid supplementation improves photosynthesis and growth in mustard through changes in proline accumulation and ethylene formation under drought stress. *South African Journal of Botany*, 98, 84-94.
63. Nishiyama, R., Watanabe, Y., Leyva-Gonzalez, M. A., Van Ha, C., Fujita, Y., Tanaka, M., and Tran, L. S. P. (2013). *Arabidopsis* AHP2, AHP3, and AHP5 histidine phosphotransfer proteins function as redundant negative regulators of drought stress response. *Proceedings of the National Academy of Sciences*, 110(12), 4840-4845.
64. Noreen, S., Ashraf, M., Hussain, M., and Jamil, A. (2009). Exogenous application of salicylic acid enhances antioxidative capacity in salt stressed sunflower (*Helianthus annuus* L.) plants. *Pak. J. Bot*, 41(1), 473-479.
65. Osman, H. E., and Fadhlallah, R. S. (2023). Impact of lead on seed germination, seedling growth, chemical composition, and forage quality of different varieties of *Sorghum*. *Journal of Umm Al-Qura University for Applied Sciences*, 9(1), 77-86.)
66. Palma, F., López-Gómez, M., Tejera, N. A., and Lluch, C. (2013). Salicylic acid improves the salinity tolerance of *Medicago sativa* in symbiosis with *Sinorhizobium meliloti* by preventing nitrogen fixation inhibition. *Plant Science*, 208, 75-82.
67. Popova, L. P., Maslenskova, L. T., Ivanova, A., and Stoinova, Z. (2012). Role of salicylic acid in alleviating heavy metal stress. *Environmental adaptations and stress tolerance of plants in the era of climate change*, 447-466.
68. Rani, M., Vikas, Kumar, R., Lathwal, M., and Kamboj, A. (2024). Effect and responses of lead toxicity in plants. In *Lead toxicity mitigation: sustainable Nexus approaches* (pp. 211-241). Cham: Springer Nature Switzerland.
69. Ruley, A. T., Sharma, N. C., Sahi, S. V., Singh, S. R., and Sajwan, K. S. (2006). Effects of lead and chelators on growth, photosynthetic activity and Pb uptake in *Sesbania drummondii* grown in soil. *Environmental pollution*, 144(1), 11-18.
70. Sadeghian, F., Hadian, J., Hadavi, M., Mohamadi, A., Ghorbanpour, M., and Ghafarzadegan, R. (2013). Effects of exogenous salicylic acid application on growth, metabolic activities and essential oil composition of *Satureja khuzistanica* Jamzad. *Journal of Medicinal Plants*, 12(47), 70-82.
71. Saleem, M., Fariduddin, Q., and Castroverde, C. D. M. (2021). Salicylic acid: A key regulator of redox signalling and plant immunity. *Plant Physiology and Biochemistry*, 168, 381-397.
72. Sharma, A., Sidhu, G. P. S., Araniti, F., Bali, A. S., Shahzad, B., Tripathi, D. K., and Landi, M. (2020). The role of salicylic acid in plants exposed to heavy metals. *Molecules*, 25(3), 540.
73. Sharma, J., Kumar, S., Kumar, V., Singh, P., Khyalia, P., Verma, S., and Sharma, A. (2023). Foliar application of glycine betaine to ameliorate lead toxicity in barley plants by modulating antioxidant enzyme activity and biochemical parameters. *Environmental Research Communications*, 5(7), 075002.

74. Sharma, P., and Dubey, R. S. (2005). Lead toxicity in plants. *Brazilian Journal of Plant Physiology*, 17(1), 35–52.
75. Sidhu, G. P. S., Singh, H. P., Batish, D. R., and Kohli, R. K. (2016). Effect of lead on oxidative status, antioxidative response and metal accumulation in *Coronopus didymus*. *Plant physiology and biochemistry*, 105, 290-296.
76. Silva, J. M., da Silva Júnior, G. B., Bonifácio, A., Dutra, A. F., de Mello Prado, R., de Alcântara Neto, F., and de Sousa, R. S. (2023). Exogenous salicylic acid alleviates water stress in watermelon plants. *Annals of Applied Biology*, 182(1), 121-130.
77. Singh, R., Hemantaranjan, A., and Patel, P. K. (2015). Salicylic acid improves salinity tolerance in field pea (*Pisum sativum* L.) by intensifying antioxidant defense system and preventing salt-induced nitrate reductase (NR) activity loss. *Legume Research-an International Journal*, 38(2), 202-208.
78. Sinha, P., Dube, B. K., Srivastava, P., and Chatterjee, C. (2006). Alteration in uptake and translocation of essential nutrients in cabbage by excess lead. *Chemosphere*, 65(4), 651-656.
79. Song, W., Shao, H., Zheng, A., Zhao, L., and Xu, Y. (2023). Advances in roles of salicylic acid in plant tolerance responses to biotic and abiotic stresses. *Plants*, 12(19), 3475.
80. Staszak, A. M., Malecka, A., Ciereszko, I., and Ratajczak, E. (2020). Differences in stress defence mechanisms in germinating seeds of *Pinus sylvestris* exposed to various lead chemical forms. *PLoS One*, 15(9), e0238448.
81. Subasinghe, C. S., Ratnayake, A. S., Roser, B., Sudesh, M., Wijewardhana, D. U., Attanayake, N., and Pitawala, J. (2022). Global distribution, genesis, exploitation, applications, production, and demand of industrial heavy minerals. *Arabian Journal of Geosciences*, 15(20), 1616.)
82. Sun, S. Q., He, M., Cao, T., Yusuyin, Y., Han, W., and Li, J. L. (2010). Antioxidative responses related to H₂O₂ depletion in *Hypnum plumaeforme* under the combined stress induced by Pb and Ni. *Environmental Monitoring and Assessment*, 163, 303-312.
83. Szepesi, Á. (2008). Influence of exogenous salicylic acid on antioxidant enzyme activities in the roots of salt stressed tomato plants. *Acta Biologica Szegediensis*, 52(1), 199-200.
84. Talha, M., Shani, M. Y., Ashraf, M. Y., De Mastro, F., Brunetti, G., Khan, M. K. R., and Coccozza, C. (2023). Lead toxicity-mediated growth and metabolic alterations at early seedling stages of maize (*Zea mays* L.). *Plants*, 12(18), 3335.
85. Vasilachi-Mitoseru, I. C., Stoleru, V., and Gavrilescu, M. (2023). Integrated assessment of pb (ii) and cu (ii) metal ion phytotoxicity on *Medicago sativa* L., *Triticum aestivum* L., and *Zea mays* L. *Plants: Insights into germination inhibition, seedling development, and ecosystem health*. *Plants*, 12(21), 3754.
86. Verma, S., and Dubey, R. S. (2003). Lead toxicity induces lipid peroxidation and alters the activities of antioxidant enzymes in growing rice plants. *Plant science*, 164(4), 645-655.

87. Wang, C., and Zhang, Q. (2017). *Exogenous salicylic acid alleviates the toxicity of chlorpyrifos in wheat plants (Triticum aestivum)*. *Ecotoxicology and environmental safety*, 137, 218-224.
88. Wang, C., Zhang, S., Wang, P., Hou, J., Qian, J., Ao, Y., and Li, L. (2011). *Salicylic acid involved in the regulation of nutrient elements uptake and oxidative stress in Vallisneria natans (Lour.) Hara under Pb stress*. *Chemosphere*, 84(1), 136-142.
89. Wang, L., Liu, B., Wang, Y., Qin, Y., Zhou, Y., and Qian, H. (2020). *Influence and interaction of iron and lead on seed germination in upland rice*. *Plant and Soil*, 455, 187-202.
90. Wang, P., Zhang, S., Wang, C., and Lu, J. (2012). *Effects of Pb on the oxidative stress and antioxidant response in a Pb bioaccumulator plant Vallisneria natans*. *Ecotoxicology and Environmental Safety*, 78, 28-34.
91. Wang, Q., Liang, X., Dong, Y., Xu, L., Zhang, X., Kong, J., and Liu, S. (2013). *Effects of exogenous salicylic acid and nitric oxide on physiological characteristics of perennial ryegrass under cadmium stress*. *Journal of plant growth regulation*, 32, 721-731.
92. Woźny, A., Zatorska, B., and Młodzianowski, F. (1982). *Influence of lead on the development of lupin seedlings and ultrastructural localization of this metal in the roots*. *Acta Societatis Botanicorum Poloniae*, 51(3-4), 345-351.
93. Wu, Z., Wang, X., Song, B., Zhao, X., Du, J., and Huang, W. (2021). *Responses of photosynthetic performance of sugar beet varieties to foliar boron spraying*. *Sugar tech*, 23, 1332-1339.
94. Xiong, Z. T., Zhao, F., and Li, M. J. (2006). *Lead toxicity in Brassica pekinensis Rupr.: effect on nitrate assimilation and growth*. *Environmental Toxicology: An International Journal*, 21(2), 147-153. Xiong, Z. T., Zhao, F., and Li, M. J. (2006). *Lead toxicity in Brassica pekinensis Rupr.: effect on nitrate assimilation and growth*. *Environmental Toxicology: An International Journal*, 21(2), 147-153.
95. Yang, H., Fang, R., Luo, L., Yang, W., Huang, Q., Yang, C., and Wang, J. (2023). *Uncovering the mechanisms of salicylic acid-mediated abiotic stress tolerance in horticultural crops*. *Frontiers in Plant Science*, 14, 1226041.
96. Zanganeh, R., Jamei, R., and Rahmani, F. (2019). *Role of salicylic acid and hydrogen sulfide in promoting lead stress tolerance and regulating free amino acid composition in Zea mays L*. *Acta Physiologiae Plantarum*, 41(6), 94.
97. Zanganeh, R., Jamei, R., and Rahmani, F. (2020). *Pre-sowing seed treatment with salicylic acid and sodium hydrosulfide confers Pb toxicity tolerance in maize (Zea mays L.)*. *Ecotoxicology and Environmental Safety*, 206, 111392.
98. Zulfiqar, F., and Ashraf, M. (2021). *Bioregulators: unlocking their potential role in regulation of the plant oxidative defense system*. *Plant Molecular Biology*, 105, 11-41.